

Computational Analysis of Bacterial Protein-Inhibitory Interactions

Presented by Julia Davies, College of Arts and Sciences, Class of 2022 Mentored by Marisa C. Kozlowski, Ph.D., Department of Chemistry, Roy and Diana Vagelos Laboratories, University of Pennsylvania, Philadelphia, Pennsylvania

Overview

- 19 Honokiol-inspired analogs shown to have inhibitory bioactivity¹ were initially docked with five potential protein targets using AutoDock Vina, a computational docking program
- The binding energy of Honokiol-inspired analogs with cell division protein FtsA were found to have a relatively strong positive correlation between Ln(MIC). Six structural variations of FtsA were further explored. Strong positive correlations emerged between the binding energies of the analogs and the FtsA proteins found in *Staphylococcus aureus*.
- Analysis of residue-ligand interactions suggested that the Honokiol-inspired analogs act as competitive inhibitors of ATP, disrupting cell division and bacterial growth.
- Insights into these antimicrobial agents mode of action informed the design of three additional analogs

Methods and Materials

Minimum Inhibitory Concentration: Minimum inhibitory concentration assays of the analogs against three representative Gram-positive bacteria (S. Epidermidis, E. Faecalis and Methicillin-Resistant S. aureus (MRSA)) was undertaken in a previous study. This data was averaged and and the natural log was calculated.

- **AutoDock Vina²:** A program in the AutoDock Suite that predicts a set of optimal bound conformations and binding energy of small molecule ligands with a target protein. The program also analyzes protein-ligand binding residues.
- Simplifications: assumes a rigid receptor and flexible ligand with flexibility specified by the user. Users must also specify search space around the receptor that the docking algorithm will explore.
- Accuracy: Binding energy is given in kcal per mol with errors of roughly 2-3 kcal/mol³.
- AutoDock Vina scores the conformations and free energies based on steric interactions:
 - Steric interactions
 - Hydrogen-bonding
 - Hydrophobicity/hydrophilicity

Computed Atlas of Surface Topography of proteins (CASTp 3.0)⁴: A web server that locates, delineates and measures the topological properties of protein structures using the alpha shape method. The server was used to identify pockets on the protein's surface as potential binding sites. Pockets with volumes greater than 50 Angstroms were considered. **PyMOL⁵:** An open-source molecular visualization system that was used to edit protein files and analyze AutoDock Vina results

Analysis: The natural log of minimum inhibitory content (Ln(MIC)) and binding energy was graphed using a simple linear regression. If the protein is the target of the ligands, then the degree of binding of the ligand to the protein should be linearly correlated to the concentration needed to inhibit bacterial growth. Small In(MIC), strong inhibition, is correlated to more negative binding energy (i.e. stronger binding).

Results

I. Minimum Inhibitory Concentration(MIC) vs Binding Energies

Graph 1. Initial Proteins Graphed by Ln(MIC) v. Binding Energy (kcal/mol)



Contact

Julia Davies BA, Chemistry, College of Arts and Sciences, University of Pennsylvania Class of 2022 Email: judavies@sas.upenn.edu





	Binding Energy of AMPPNP Form of S. aureus	
Analog	(kcal/mol)	Binding Energy of ATP Form of <i>S. aureus</i> (kcal/mol)
CRO215	-8.8	-8
CRO254	-8.9	-9
CRO209	-9.0	-9
CRO312	-9.1	-9
CRO262	-9.3	-9
HFR198	-9.4	-9
HFR199	-9.6	-9
Analog 1	-9.3	-9.
Analog 2	-9.3	-9.
Analog 3	-9.7	-9.
ATP	-10	-10

II. Structure of Best Analogs and Design of New Analogs

Figure 2.

Dhanal Drataction	Linker Group	jun Opri
Phenol Protection		0
None	OH CRO215 OH	OH OH HFR198 OH
یم مرب	CRO209 OH	HFR199 OH
22	O CRO254	Analog 1 OH
n de la constante de la consta	CRO312 OH	Analog 2 OH
m	CRO262	O O O O H Analog 3



•	A stro dockii neces
•	of ATF Six ot in the
•	energ <i>Staph</i> condit
•	hospit inhibit Furthe
	inspire comp inhibit
•	conce The a used
	Staph comp comp

References

- 1. Solinski, A.; Ochoa, C.; Lee, Y.; Paniak, T.; Kozlowski, M.; Wuest, W. Honokiol-Inspired Analogs As Inhibitors Of Oral Bacteria. ACS Infectious Diseases 2017, 4, 118-122. 2. Trott, O.; Olsen, A. AutoDock Vina: improving the speed and accuracy of docking with a new scoring function, efficient optimization and multithreading. J Comput. Chem. 2010, 31(2) 455-461
- 4. Tian et al., Nucleic Acids Res. 2018
- 5. The **PyMOL** Molecular Graphics System, Version 1.2r3pre, Schrödinger, LLC.
- 6. Chick, H. An investigation into the laws of disinfection. Journal of Hygiene (Cambridge) 1908 8 92-158. 1879-1885.
- 8. Loose, M.; Mitchison, T. The bacterial cell division proteins FtsA and FtsZ self-organize into dynamic cytoskeletal patterns. Nat Cell Biol. 2014; 16(1) 38-46.



IV. Residue-Ligand Interactions

Discussion

ong positive correlation was found between the analogs' In(MIC) and binding energy resulting from the ing with cell-division protein FtsA expressed in *Staphylococcus aureus* subsp. aureus MW2. FtsA is ssary for cell division and inhibition of this protein leads to filamentation and eventual cell death. Binding P to FtsA is required for its proper functioning⁸.

ther versions of FtsA expressed in both Staphylococcus aureus and Thermotoga maritima were analyzed same way with strong positive correlations found between the analogs' average ln(MIC) and binding y with FtsA as it is expressed in *Staphylococcus aureus*.

hylococcus aureus subsp. MRSA252 is a gram-positive bacteria that grows in anaerobic and aerobic itions. It is a methicillin-resistant strain of Staphylococcus aureus and is a major cause of community- and tal- acquired infections. This form of FtsA most likely matches the bacteria screened in the minimum tory concentration assays which included MRSA.

er analysis of ligand-residue interactions on AutoDock Vina and PyMol showed that the 19 Honokiolred analogs were specifically binding in the ATP (or AMPPNP) binding spot of FtsA and may be acting as petitive inhibitors to ATP and inhibiting cell division. Although these analogs may be acting as competitive itors to ATP, ATP consistently had a lower binding energy with FtsA which indicates that a higher entration of the analogs compared to ATP is required for an inhibitory effect.

analogs with the lowest predicted binding energy were identified and the features of these analogs were to design three new analogs. Initial docking of these three newly designed compounds with hylococcus aureus subsp. MRSA252 FtsA revealed lower binding energies compared to the parent bound CRO215, with Analog 3 having among the lowest predicted binding energies among all the bounds, suggesting that it may be an effective antibacterial agent.

Conclusions

This research explores the cell-division protein FtsA as a potential target for antibacterial agents and potential FtsA inhibitors.

Honokiol-inspired analogs which have shown to have inhibitory bioactivity may be acting at competitive inhibitors to ATP binding with the cell-division protein FtsA.

Three new analogs, including 4-(*tert*-butyl)-3-hydroxyphenyl 4-(*tert*-butyl)-3-(prop-2-yn-1loxy)benzoate, are proposed and preliminary predicted binding energy suggests that these

analogs may work as antibacterial agents. Further biological experiments should be conducted to explore the effectiveness of these analogs as antibacterial agents.

Further research should be conducted into the role of FtsA in cytokinesis as FtsA inhibitors may be appealing antibacterial agents.

Acknowledgements

Research Supervisors: Dr. Marisa Kozlowski and Hanna Roenfanz Funding Source: Summer 2020 Penn Undergraduate Research Program (PURM) Special thanks to Hanna Roenfanz conducting biological assays and sharing MIC data

3. Forli, S.; Huey, R.; Pique, M.; Sanner, M.; Goodsell, D.; Olsen, A. Computational protein-ligand docking and virtual drug screening with the Autodock Suite. Nature Protocols. 2016, 11(5), 905-919.

7. Fujita, J.; Maeda, Y.; Nagao, C.; Tsuchiya, Y.; Miyazaki, Y.; Hirose, M.; Matsumura, H.; Mizohata, E.; Matsumoto, Y.; Inoue, T.; MIzuguchi, K.; Matsumura, H. Crystal structure of FtsA from Staphylococcus aureus. FEBS letters, 2014; 588(10),