

High Throughput Identification of Developmental Regulators in Early *C. elegans* Embryos

Perelman School of Medicine UNIVERSITY of PENNSYLVANIA

William "Jack" Marchese CAS 2024 Email: marchwil@sas.upenn.edu

Advised by Dr. John Isaac Murray, Elicia Preston, and Dr. Jonathan Rumley of the Department of Genetics

INTRODUCTION

The objective of this project was to create and image strains of *C. elegans* with markers on previously unexplored early developmental genes. The genes of interest included the *tbx-31*, *tbx-32*, *tbx-33*, and *tbx-39* *tbx* genes, as well as the *ceh-32* and *ceh-76* homeotic genes. Using state of the art imaging software, the expression of these genes was tracked in individual cells of developing embryos from the 2 cell stage to the larval stage of young *C. elegans*.

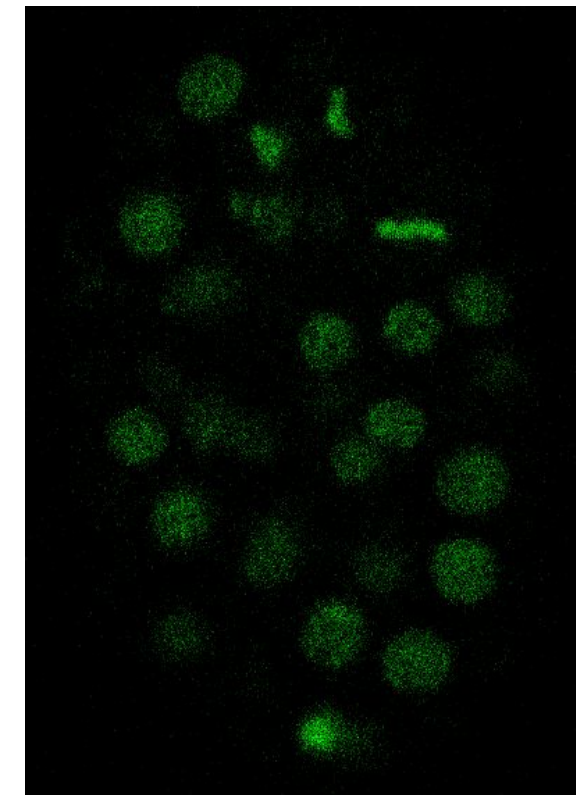
METHODS

The new strains were created by crossing existing strains with the requisite RFP marker on the gene of interest into a strain with GFP histone lineaging markers in order to track cell expression in developing embryos. Using ACETREE imaging software, the expression of RFP tagged genes was tracked over time. This was used to develop an ancestral tree of cell lineages, highlighting the individual cells in which the genes of interest were expressed.

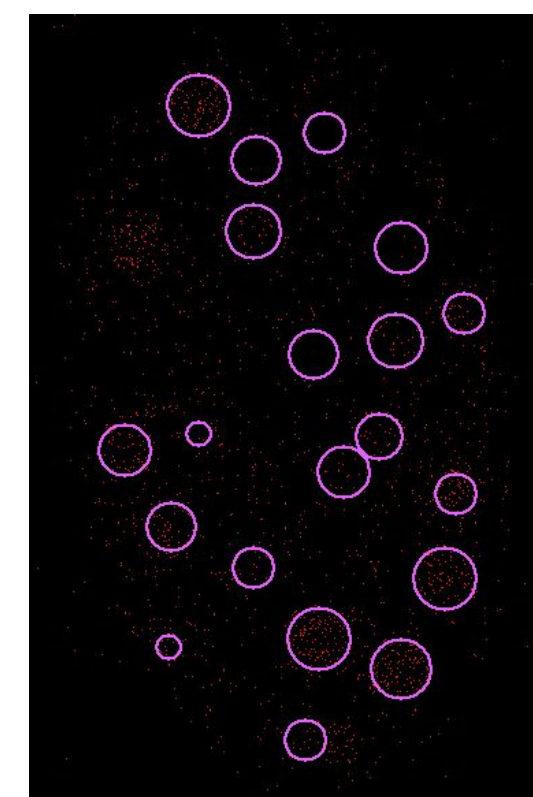
RESULTS

Viable strains were created for the *tbx-31*, *tbx-32*, *tbx-39*, and *ceh-32* genes through multiple rounds of backcrossing the RFP-tagged genes of interest into a GFP lineage reporter strain.

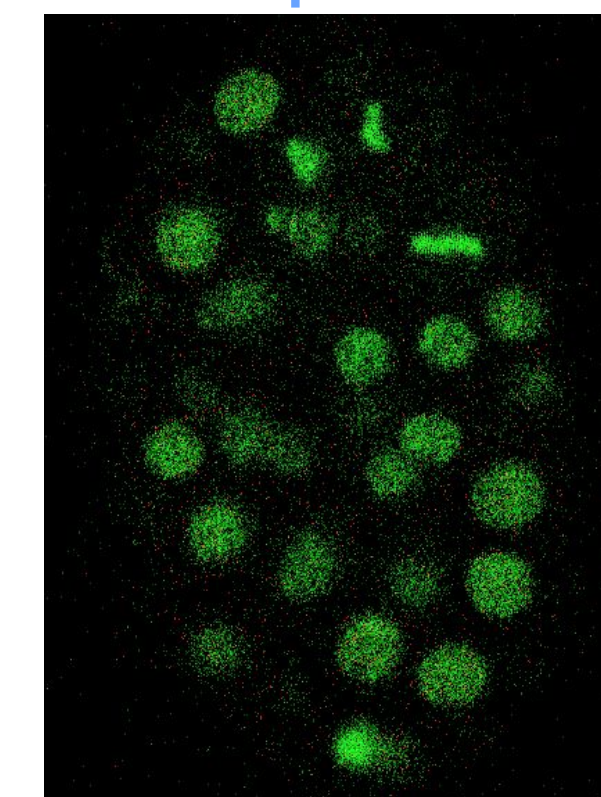
GFP lineage reporter



RFP *tbx-39* reporter

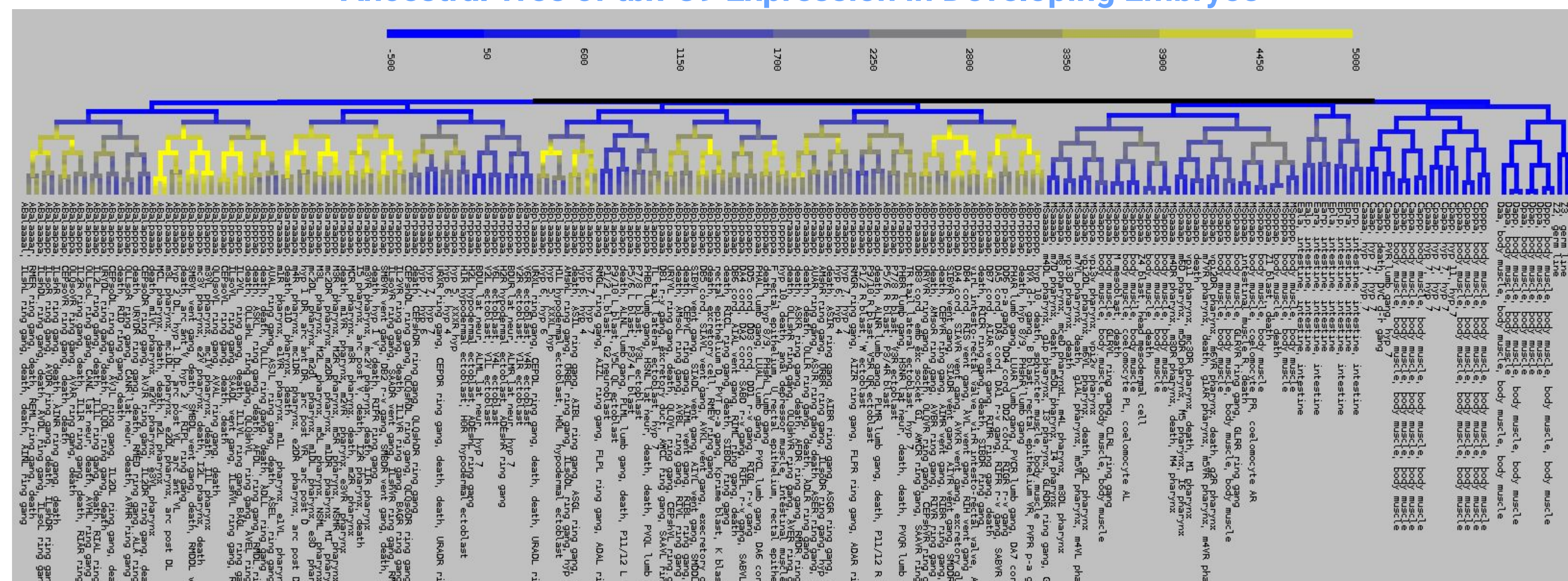


Combined Expression



Pictured above is the expression data for a *tbx-39* embryo. The green lineage marker tracks all cell positions over time. The red marker shows where *tbx-39* was expressed in the developing embryo.

Ancestral Tree of *tbx-39* Expression in Developing Embryos



The ancestral tree below shows the results of the *tbx-39* imaging. Cells highlighted in yellow showed *tbx-39* expression. The *tbx-39* gene appears to be expressed exclusively in the AB cell lineage. It appears that while *tbx-39* is not tissue specific, the plurality of cells in which it is expressed become pharyngeal cells.

CONCLUSION

The development of these new worm strains for the imaging of early developmental regulators elucidates the temporal and spatial expression of several early *tbx* and homeotic genes. Specifically, the data gathered on *tbx-39* opens the door for further research into its interactions with other genes, in the hopes of understanding new signaling pathways in development.

FUTURE RESEARCH

The other newly developed strains with RFP tagged *tbx* and homeotic genes will need to be imaged and an ancestral trees of gene expression will be determined. Additionally, mutants for these genes are in the process of being developed through CRISPR knockouts, which, combined with ACETREE, will help determine the function of these early developmental regulators.

ACKNOWLEDGEMENTS

Many thanks to PURM and all of CURF for this research opportunity, as well as Dr. John Isaac Murray, Elicia Preston, and Dr. Jonathan Rumley of the Murray lab within the Perelman School of Medicine for guiding me this summer