

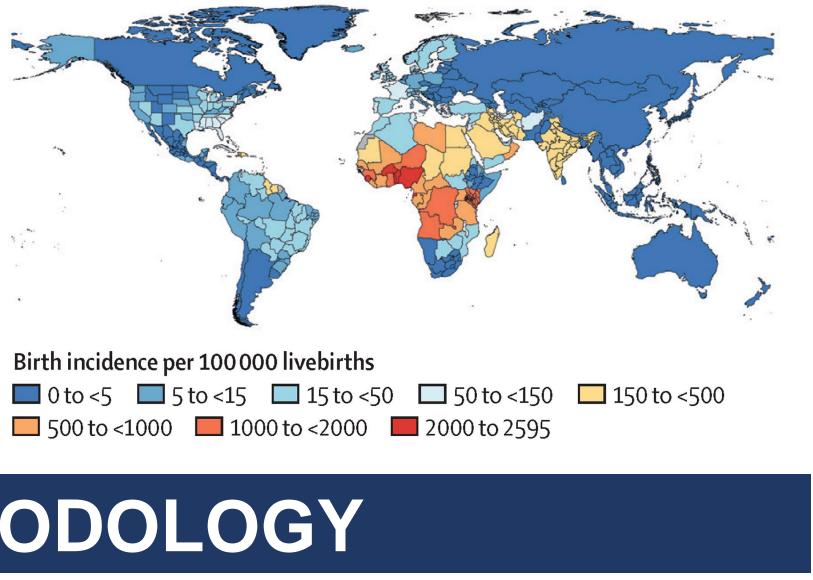
ABSTRACT

Although thousands of people worldwide suffer from sickle cell disease, the most common form of an inherited blood disorder, there are only a handful of available treatments available to combat the significant morbidities, lifelong challenges, reduced quality of life, and early mortality rates in patients. The resources available are also inaccessible to many either due to price or the possibility of adverse effects such as myelosuppression. Thus, to discover more effective and affordable treatments for patients with sickle cell disease, we investigated the antisickling effect of SCD-101, a botanical extract often utilized for medicinal purposes in Nigeria and other West African countries. After conducting multiple in-vitro sickling assays, we discovered that there are one or more small molecules located in fractions of SCD-101 that confer the indigenous plant with anti-sickling activity. In view of the outcome of this study, SCD-101 may be a promising option for the treatment of patients with sickle cell disease.

BACKGROUND

Sickle cell disease (SCD) is an inherited genetic disorder that affects approximately 100,000 Americans and 20 - 25 million people worldwide. Caused by a singular substitution of value for glutamic acid in the sixth position of the β -chain of hemoglobin S (HbS), SCD is a debilitating blood disorder that impairs an individual's ability to live a healthy, pain-free life. The pathophysiology of SCD is based on polymerization of deoxygenated sickle hemoglobin (HbS), which leads to red blood cell (RBC) sickling. This phenomenon occurs under conditions of low oxygen (O_2) saturation known as hypoxia. The consequence of RBC sickling is a systemic illness that manifests primarily as painful crises, leading to progressive damage, poor quality of life and a decreased life expectancy in many organ patients. Although two gene therapy approaches for complete cure of SCD were approved in the US recently, novel therapies that are both effective and cheap to manufacture remain a top priority because the primary disease burden is in the developing world, specifically, in sub-

Saharan Africa where the most affected individuals live. This unfortunate reality provides the opportunity for, and justifies the importance of my project, which seeks to investigate SCD-101, a botanical extract from the Sorghum bicolor plant grown in Nigeria. Although its mechanism of action and active substance composition remain unknown, evidence suggests that ≥ 1 small molecules are responsible for its activity.



METHODOLOGY

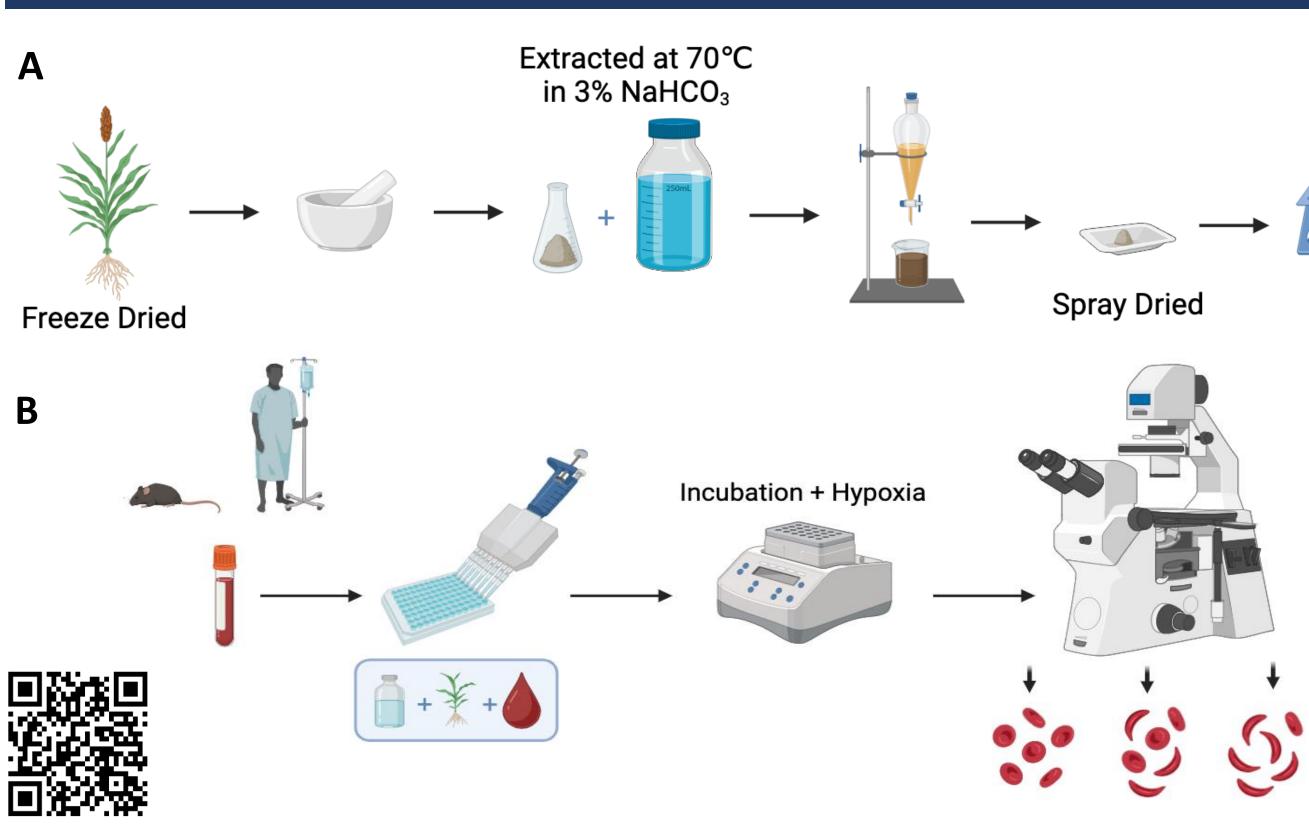
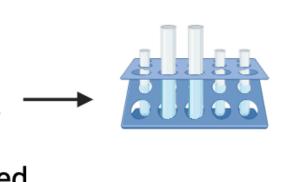


Figure 1: (A) Methodology: SCD-101 Fraction Extraction **(B)** Methodology: Sickling Assay Protocol

In vitro Studies on SCD-101, A Promising Botanical **Extract with Anti-sickling Properties for Sickle Cell Disease Therapy**

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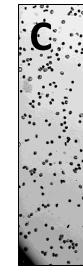
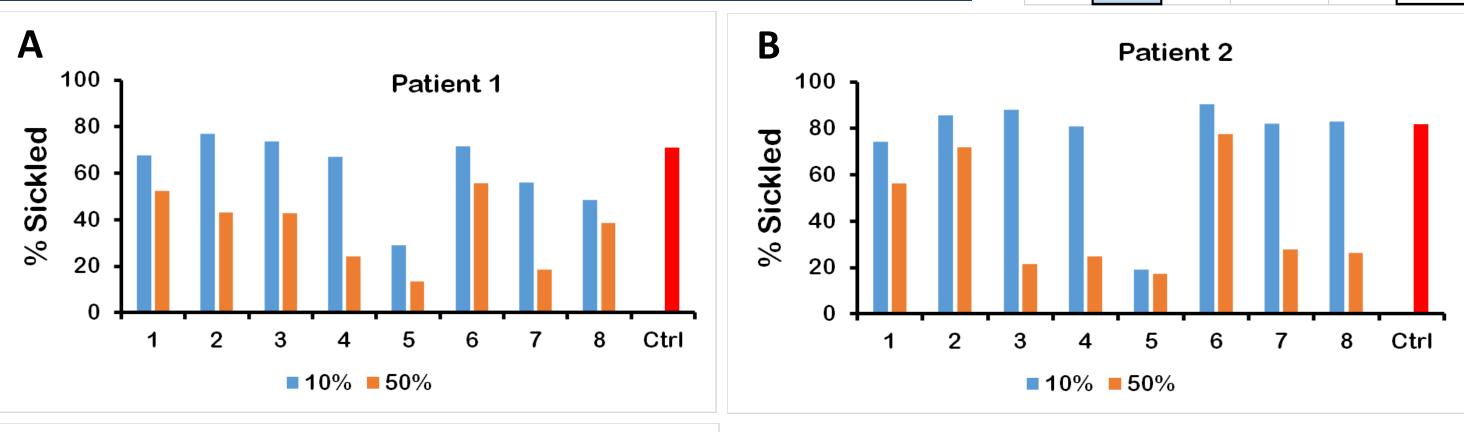
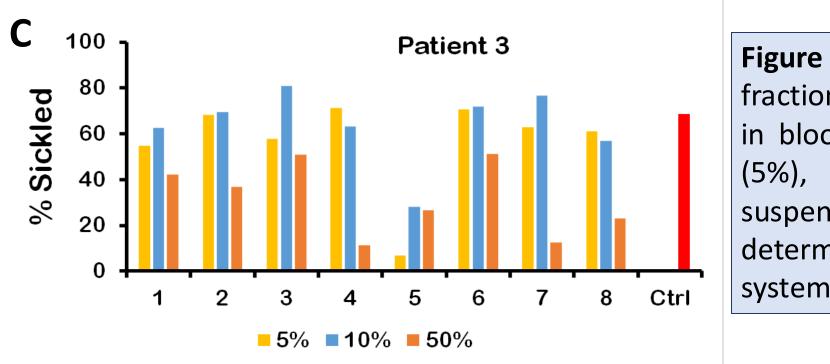


Figure 2: Representative image of (A) healthy red blood cells treated with 5% of fraction 5 post incubation and 1 hour hypoxia (B) untreated, sickled red blood cells after 1 hour hypoxia (C) red blood cells with spherocytosis (**D**) red blood cells with echinocytosis.

			<u> </u>	AU U				Samula	Imaga	Sickled Cells			
	Sample	Image	Sickled Cells	All cells	% Sickled			Sample	Image		-		
Patient 1	L1	25	332	490	67.7551		Patient 2	L1	41	262	L		
	L2	26	409	531	77.02448				L2	42	723		
	L3	27	474	644	73.60248				L3	43	817		
	L4	28	370	551	67.15064				L4	44	573		
	L5	29	73	253	28.85375				L5	45	174		
	L6	30	449	627	71.61085			L6	46	761			
	L7	31	601	1072	56.06343			ent	L7	47	650		
	L8	32	179	370	48.37838				<u> </u>	L8	48	563	
	H1	33	323	618	52.26537			H1	49	241			
	H2	34	239	554	43.14079			at	H2	50	511		
	H3	35	295	690	42.75362			H3	51	112			
	H4	36	242	995	24.32161				_	H4	52	124	
	H5	37	2	15	13.33333				H5	53	60		
	H6	38	448	807	55.51425					H6	54	519	
	H7	39	112	603	18.5738				H7	55	160		
	H8	40	242	629	38.47377				H8	56	165		
	Ctrl				70.86331			Ctrl					

Figure 3: Left) Results obtained using blood samples from Patient 1 Middle) Results obtained using blood samples from Patient 2 Right) Results obtained using blood samples from Patient 3





SECONDARY DATA

	Sample	Image	Sickled cells	All cells	% Sickled				
5%	Fraction 5	1	104	500	20.8				
	Fraction 5 (pH)	2	295	603	48.92206		100 •	_	
	Fraction 1	3	493	732	67.34973		100 .	1	
	Fraction 1 (pH)	4	363	628	57.80255	σ	80 ·	4	
	Fraction 5	5	168	575	29.21739	Sickled	~~		
100/	Fraction 5 (pH)	6	174	526	33.07985	×	60 ·	1	
10%	Fraction 1	7	509	695	73.23741	<u>Sic</u>	40 •	4	
	Fraction 1 (pH)	8	281	434	64.74654				
	Fraction 5	9	100	408	24.5098	%	20 •	1 🗖	
E 00/	Fraction 5 (pH)	10	52	379	13.72032		0 -		
50%	Fraction 1	11	281	393	71.50127		0	' Fra	
	Fraction 1 (pH)	12	324	457	70.89716			га	5
Positive	502 (1.5 mM)	13	112	523	21.41491				3
Controls	VOX (0.75 mM)	14	82	416	19.71154				
Negative	Ctrl 1	15	341	567	60.14109				
Controls	Ctrl 2	16	301	516	58.33333				

Figure 5: Left) Results collected from sickling assay performed using blood samples from a patient with SCD. Fraction 5, Fraction 1, Fraction 5 (pH = 7.4), Fraction 1 (pH = 7.4), positive and negative controls were tested. **Right)** Bar graph that depicts the effect of various suspensions on the % of sickled cells after 1 hour hypoxia.

PRELIMINARY DATA

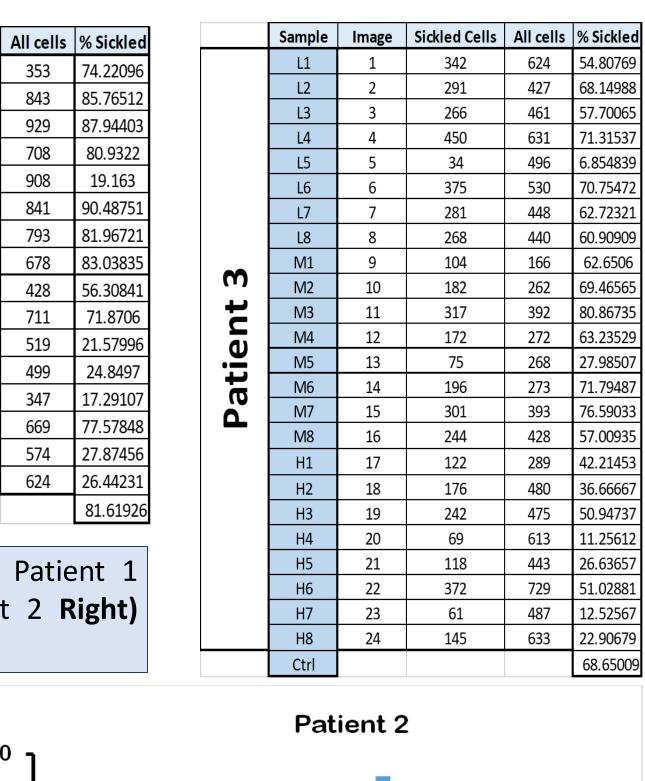
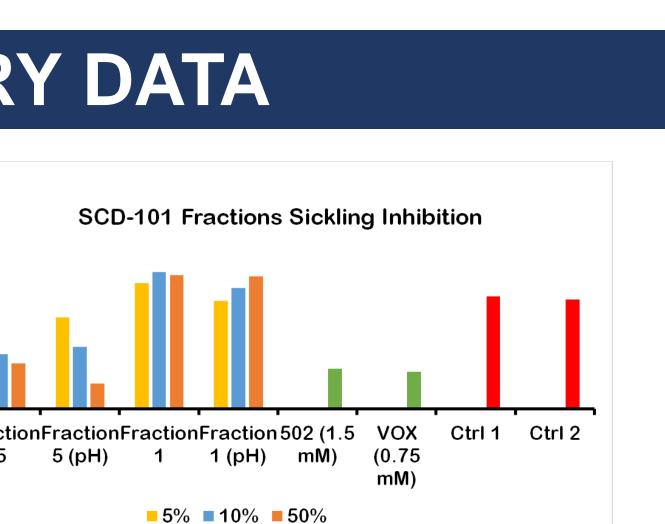


Figure 4: Bar graphs that depicts the effect of various fractions of SCD-101 on the percentage of sickled cells in blood sample from three different patients at low (5%), intermediate (10%), and high (50%) vol/vol suspensions. The percentage of sickled cells was determined using a computer-assisted image analysis system called ImageJ and graphed using Excel.



CONCLUSION

- and technical replicates)
- by at least 50%.

If we can definitively identify the fractions that contain one or more small molecules responsible for inhibiting the polymerization of HbS, then we can subsequently concentrate the active components to improve the potency of the extract as well as the dosage requirement if it were to be developed into a drug. In addition, gaining a better understanding about the plant's activity may enable us to learn more about how its anti-sickling agents interact with hemoglobin in vitro and in vivo.

The main implication from this experiment is the potential of botanical plants in the treatment of SCD. Botanical plants have long been utilized as a source of medicine by various communities worldwide. Particularly, in developing countries, medicinal plants are heavily relied upon due to their cultural significance as well as accessibility. Unable to afford the cost of treatment offered by traditional hospitals and medical professions, many individuals manage diseases, such as SCD by using crude extracts from plants. Thus, I believe that the investigation of botanical plants could lead to various scientific advances in the drug development industry

ACKNOWLEDGMENTS

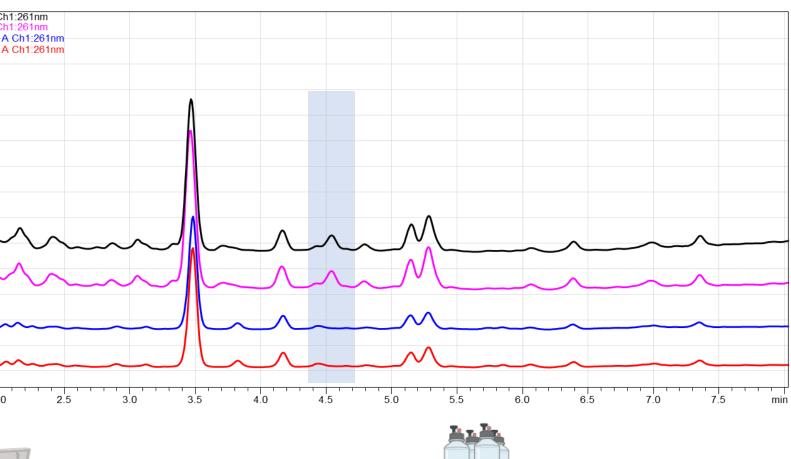
Thank you to Dr. Osheiza Abdulmalik, DVM, for sponsoring this work, guiding me throughout the process and expanding my knowledge on the drug discovery industry for SCD. In addition, thank you to Dr. Robert Swift, PhD, for providing me with the SCD-101 fractions necessary to complete multiple in-vitro sickling assays.

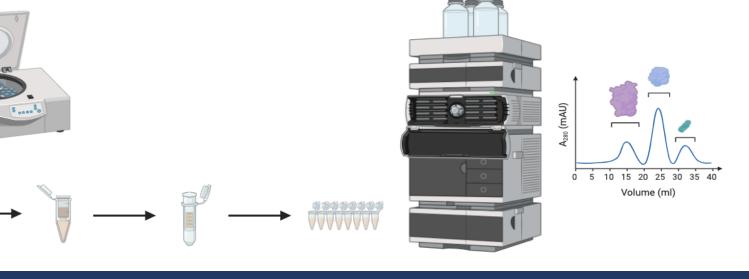


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FRACTION 5 INVESTIGATION





• The results of the sickling assay indicate that there are one or more small molecules found in fraction 5 that are responsible for the extract's anti-sickling effects.

• Fraction 5's activity was conserved across blood samples from multiple patients (biological

• Fraction 5 significantly reduced the percentage of sickled cells at both high and low volumes

• Although other fractions like fraction 7 showed some promising anti-sickling activity, albeit at high volumes, the morphology of the RBCs was compromised.

• The results of the sickling assay indicate that fraction 5 is the most reliable candidate for further fractionation and investigation as the primary anti-sickling component of SCD-101.

FUTURE DIRECTIONS

MAIN IMPLICATIONS

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